

09/373, 984

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=> s amplif##### RNA or synthes##### RNA

L1 5546 AMPLIF##### RNA OR SYNTHES##### RNA

=> s l1 and ((sequential(10a) add#####)or(serial(10a)add#####))

L2 0 L1 AND ((SEQUENTIAL(10A) ADD#####) OR(SERIAL(10A) ADD#####))

=> s l1 and step wise

L3 1 L1 AND STEP WISE

=> s l3 and (different buffer# or different reagent#)

L4 0 L3 AND (DIFFERENT BUFFER# OR DIFFERENT REAGENT#)

=> d l3 bib ab kwic

L3 ANSWER 1 OF 1 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

AN 1988:418278 BIOSIS

DN BA86:80890

TI CHANGES IN ACTIVATION MARKERS AND CELL MEMBRANE RECEPTORS ON HUMAN PERIPHERAL BLOOD T LYMPHOCYTES DURING CELL CYCLE PROGRESSION AFTER PHA STIMULATION.

AU POULTON T A; GALLAGHER A; POTTS R C; BECK J S

CS DEP. PATHOL., UNIV. DUNDEE, NINEWELLS HOSP. AND MED. SCH., DUNDEE DD2 1UB, U.K.

SO IMMUNOLOGY, (1988) 64 (3), 419-426.

CODEN: IMMUAM. ISSN: 0019-2805.

FS BA; OLD

LA English

AB Phytohaemagglutinin (PHA)-stimulated peripheral blood lymphocytes were examined sequentially for changes in volume, the appearance of cell membrane receptors and nucleic acid synthesis. The kinetics of appearance of activation antigens were compared with the progress of the cell through the separate events of volume growth and nucleic acid syntheses, to determine points at which regulation of receptors may control further progress through the cell cycle. In all samples tested there was a consistent pattern of response in the proportion of cells progressing through the cell cycle. Most of the T cells increased in size (mean 82% at 24 hr), fewer cells entered the G1a/G1b phase with the onset of RNA synthesis (mean 68% at 48 hr) and even fewer entered DNA synthesis (mean 42% at 72 hr). The time-course of appearance and the number of cells expressing IL-2 receptors were almost identical with that of cells responding by RNA synthesis. A similar correlation was observed between expression of the transferrin receptor and DNA synthesis. Addition of

anti-Tac antibody temporarily suppressed the onset of RNA synthesis and antibodies to the transferrin receptor suppressed DNA synthesis. These linkages are further evidence that IL-2 and transferrin are the specific signals for cellular RNA and DNA synthesis. With optimal concentrations of PHA, addition of IL-2 did not increase the proportion of cells bearing activation antigens or undergoing nucleic acid synthesis. Suboptimal concentrations of PHA produced a small reduction in the number of cells expressing the IL-2 receptor, but a much greater reduction in the rate of entry into RNA synthesis. There was a consistent increase in all activation parameters tested with the addition of IL-2, but the proportion of cells expressing the transferrin receptor and entering DNA synthesis was consistently lower than that of cells that expressed the IL-2 receptor or entered RNA synthesis. This suggests that regulation of the IL-2 receptor is not responsible for the reduction in the number of cells that proceed to proliferation. The CD2 antigen (T111) showed increasing expression in a **step-wise** fashion after activation, the increases coinciding with the onset of RNA and DNA syntheses.

AB. . . the reduction in the number of cells that proceed to proliferation. The CD2 antigen (T111) showed increasing expression in a **step-wise** fashion after activation, the increases coinciding with the onset of RNA and DNA syntheses.

IT Miscellaneous Descriptors

PHYTOHEMAGGLUTININ NUCLEIC ACID **SYNTHESIS** RNA DNA
INTERLEUKIN 2 TRANSFERRIN

=>

WEST**Freeform Search**

09/373984

Database:

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US Pre-Grant Publication Full-Text Database
JPO Abstracts Database
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Derwent World Patents Index
IBM Technical Disclosure Bulletins

Term:

L3 and (different buffer\$1 or different reagent\$1)

Display:

10

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result set

DB=USPT,JPAB,EPAB,DWPI; PLUR=YES; OP=ADJ

<u>L4</u>	L3 and (different buffer\$1 or different reagent\$1)	5	<u>L4</u>
<u>L3</u>	L2 and ((sequential near5 add\$5) or (serial near5 add\$5))	158	<u>L3</u>
<u>L2</u>	amplif\$7 RNA or synthes\$5 RNA	3577	<u>L2</u>
<u>L1</u>	6582906.pn.	2	<u>L1</u>

END OF SEARCH HISTORY

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Search Results - Record(s) 1 through 5 of 5 returned.

-
- ☐ 1. 6551784. 09 May 01; 22 Apr 03. Method of comparing nucleic acid sequences. Fodor; Stephen P. A., et al. 435/6; 435/283.1 435/287.1 435/288.3 435/288.7 536/23.1 536/24.3. C12Q001/68 C12M001/34 G01N033/50 C07H021/04 C07H021/02.
-
- ☐ 2. 6309822. 23 Dec 96; 30 Oct 01. Method for comparing copy number of nucleic acid sequences. Fodor; Stephen P. A., et al. 435/6; 435/287.2 435/288.3 435/288.7 536/23.1 536/24.3 536/24.31. C12Q001/68 C12M001/34 C07H021/04 C07H021/02.
-
- ☒ 3. 6197557. 10 Sep 98; 06 Mar 01. Compositions and methods for analysis of nucleic acids. Makarov; Vladimir L., et al. 435/91.2; 435/6 536/23.1 536/24.3. C12P019/34 C12Q001/68 C07H021/02 C07H021/04.
-
- ☒ 4. 6190691. 15 Jun 98; 20 Feb 01. Methods for treating inflammatory conditions. Mak; Vivien H. W.. 424/449; 514/859 514/861 514/863 514/886 514/887 604/20. A61F013/00.
-
- ☐ 5. 5962477. 15 Jun 98; 05 Oct 99. Screening methods for cytokine inhibitors. Mak; Vivian. 514/327; 424/78.05. A61K031/445.
-

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Term	Documents
DIFFERENT	2694524
DIFFERENTS	197
BUFFER\$1	0
BUFFER	493686
BUFFERA	10
BUFFERB	5
BUFFERC	2
BUFFERD	45
BUFFERE	53
BUFFERF	2
BUFFERG	1
(L3 AND (DIFFERENT BUFFER\$1 OR DIFFERENT REAGENT\$1)).USPT,JPAB,EPAB,DWPI.	5

There are more results than shown above. [Click here to view the entire set.](#)

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L4: Entry 4 of 5

File: USPT

Feb 20, 2001

DOCUMENT-IDENTIFIER: US 6190691 B1

TITLE: Methods for treating inflammatory conditions

Detailed Description Text (163):

The CTLL assay for IL-2 (T-cell Growth Factor, or TCGF) can be used to quantitate the amount of IL-2 present in samples taken from cells incubated with putative anti-inflammatory agents (see, e.g., Gills, S., et al., J. Immunol. 120:2027 (1978)). Murine (C57BL/6) tumor-specific cytotoxic T lymphocytes (CTLL 1 and CTLL 2, described by Gills, et al., Nature, 268:154 (1977) and J. Exp. Med. 146:168 (1977)) are maintained in a IL-2-dependent long-term proliferative culture. The cells are washed free of growth medium using standard techniques and resuspended in Click's medium (Altich Assoc., Hudson, Wash.) supplemented with 2% FCS, 50 units/mL penicillin, 50 .mu.g/mL gentamicin, 16 .mu.g/mL NaHCO₃ and 25 .mu.M/mL HEPES and placed in microtiter wells. The supernatant to be assayed is then added to the cells in serial dilutions. The plates are incubated at 37.degree. C. in a humidified atmosphere of 5% CO₂ in air. Following 24 hours of culture, 0.5 .mu.Ci of ³H-thymidine (Schwartz/Mann Inc., Orangeburg, N.J.) having a specific activity of 1.9 .mu.Ci/mM is added to each well and the culture continued for another 4 hours. The cultures are harvested onto glass fiber filter strips and the ³H-thymidine incorporation is determined using known techniques (Oppenheim, et al., MANUAL OF CLINICAL IMMUNOLOGY, American Society for Microbiology, Washington, D.C., Rose, Ed. (1976), which is incorporated herein by reference).

Detailed Description Text (183):

In order to perform PCR amplification of RNA, it must first be transcribed into cDNA using a reverse transcriptase enzyme. Many different protocols exist for this reaction, but one that is used commonly is as follows. A solution is heated to 65.degree. C. for 5 minutes to eliminate any secondary structure of the RNA and then immediately frozen by placing on dry ice or by putting in a -70.degree. C. freezer. A mixture is then added to the RNA such that the final concentration of reagents is 20 U/.mu.l Moloney murine leukemia virus reverse transcriptase (such as that available from GIBCOBRL), 1X buffer (which is supplied with the enzyme by the manufacturer), 1.3 U/.mu.l RNase inhibitor, 50 .mu.g/mL oligo dT, 0.1 mg/mL acetylated bovine serum albumin, 0.5 mM dNTP (a mixture of dATP, dCTP, dGTP, and dTTP each at a concentration of 0.5 mM), and 10 mM dithiothreitol. The reaction mixture is incubated at 42.degree. C. for 1 hour and subsequently at 95.degree. C. for 5 minutes to inactivate the enzymes. This reaction results in the generation of cDNA species for only the mRNA fraction of the total RNA due to the use of an oligo dT primer, which binds to the poly A tail of eukaryotic mRNA. Other species of RNA as well as mRNA can be transcribed into cDNA by using random hexamer oligodeoxyribonucleotide primers. Polymerizing agents suitable for synthesizing a complementary, copy-DNA (cDNA) sequence from the RNA template are reverse transcriptase (RT), such as avian myeloblastosis virus RT, Moloney murine leukemia virus RT, or Thermus thermophilus (Tth) DNA polymerase, a thermostable DNA polymerase with reverse transcriptase activity marketed by Perkin Elmer Cetus, Inc. Typically, the RNA template is heat degraded during the first denaturation step after the initial reverse transcription step leaving only DNA template.

Detailed Description Text (188):

The PCR method can be performed in a step-wise fashion, where after each step new reagents are added, or in a fashion where all of the reagents are added simultaneously, or in a partial step-wise fashion, where fresh or different reagents are added after a given number of steps. For example, if strand separation is induced by heat, and the polymerase is heat-sensitive, then the polymerase must be replenished following each round of strand separation. However, if, for example, a helicase is used for denaturation, or if a thermostable polymerase is used for extension, then all

of the reagents may be added initially, or, alternatively, if molar ratios of reagents are of consequence to the reaction, the reagents may be replenished periodically as they are depleted by the synthetic reaction.